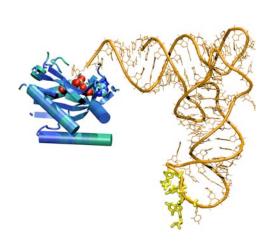
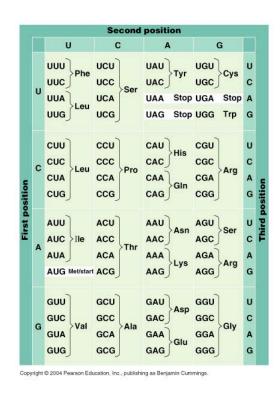
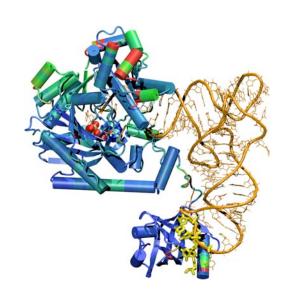
MULTISEQ in VMD -

Revealing How Nature Designs Proteins and RNAs



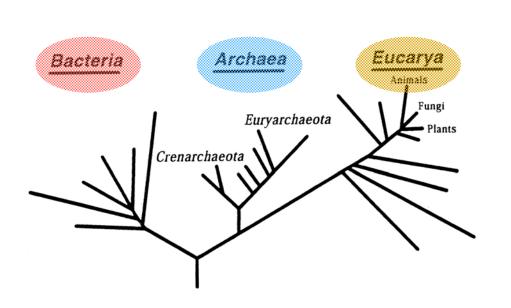


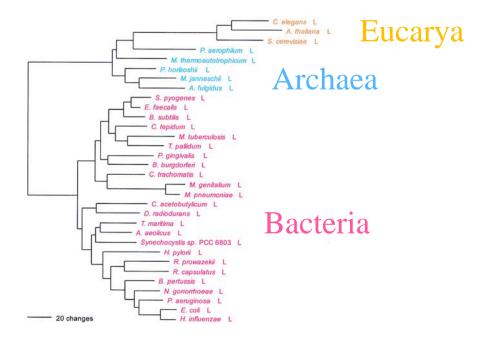


Luthey-Schulten Group

Department of Chemistry, Biophysics, and Beckman Institute University of Illinois at Urbana-Champaign

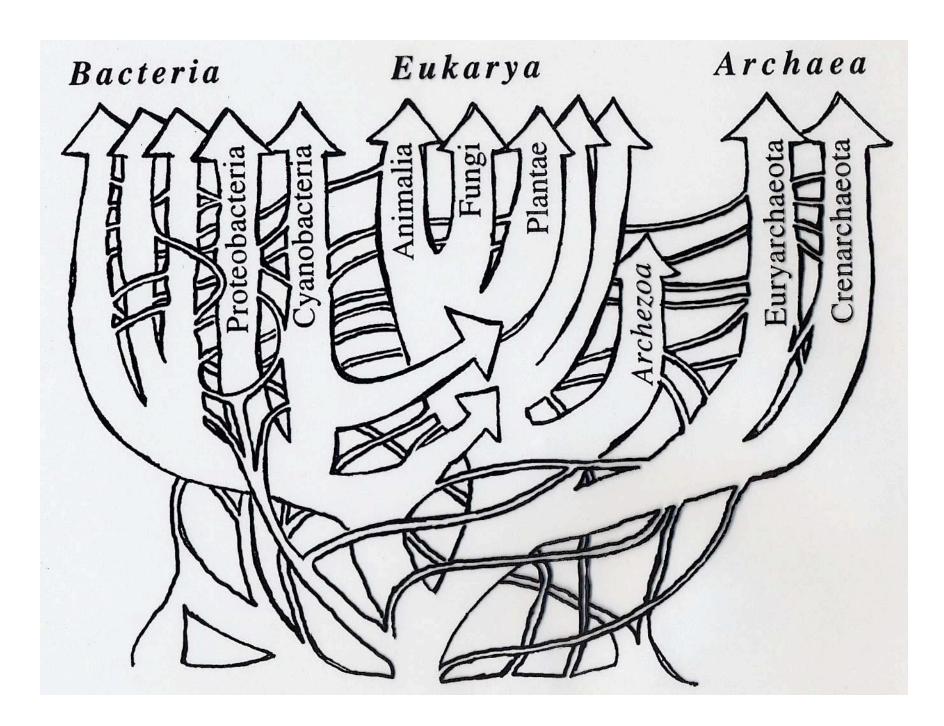
Universal Phylogenetic Tree three domains of life





Based on 16S rRNA

Leucyl-tRNA synthetase displays the full canonical phylogenetic distribution.



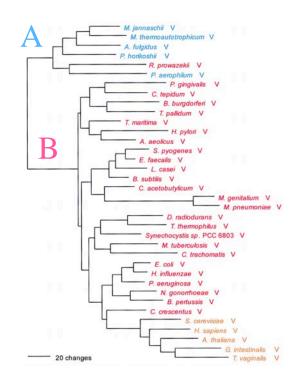
After W. Doolittle, modified by G. Olsen

Phylogenetic Distributions

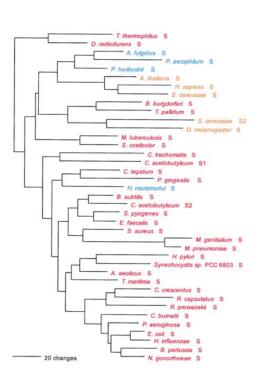
Full Canonical

A. thaliana L. S. cerevisiae L P. horikoshii L. - M. jannaschii I E. faecalis L B. subtilis L C. tepidum L - M. tuberculosis L T. pallidum L P. gingivalis L B. burgdorferi L C. trachomatis L - M. genitalium L - M. pneumoniae L C. acetobutylicum L D. radiodurans L T. maritima L A aeolicus L Synechocystis sp. PCC 6803 L H. pylorii L - R. prowazekii R. capsulatus L B. pertussis L N. gononhoeae L. P. aeruginosa L - E. coli L - 20 changes

Basal Canonical



Non-canonical



increasing inter-domain of life Horizontal Gene Transfer

"HGT erodes the historical trace, but does not completely erase it...." G. Olsen

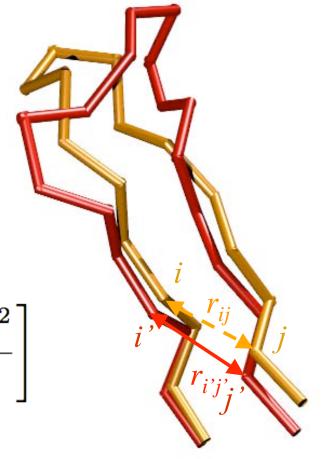
Protein Structure Similarity Measure

Q_H Structural Homology

fraction of native contacts for aligned residues + presence and perturbation of gaps

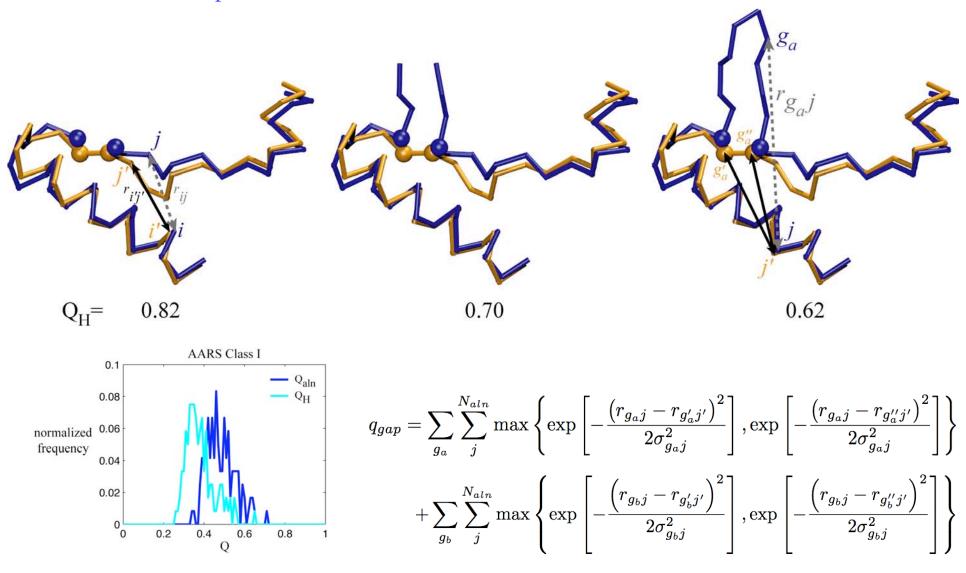
$$Q_H = \aleph \left[q_{aln} + q_{gap} \right]$$

$$q_{aln} = \sum_{i < j-2} \exp \left[-\frac{(r_{ij} - r_{i'j'})^2}{2\sigma_{ij}^2} \right]$$

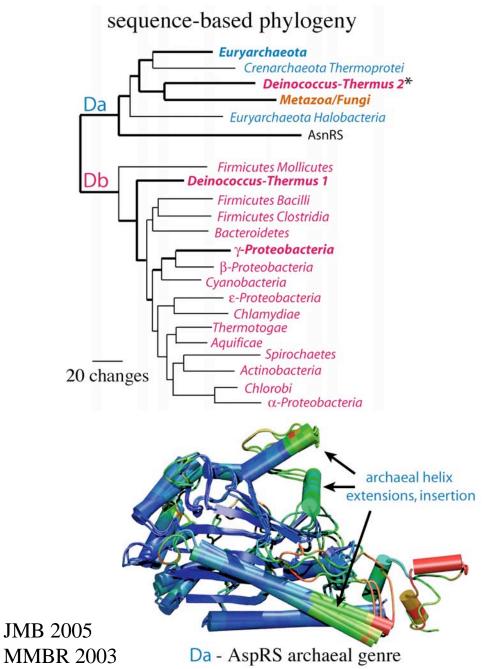


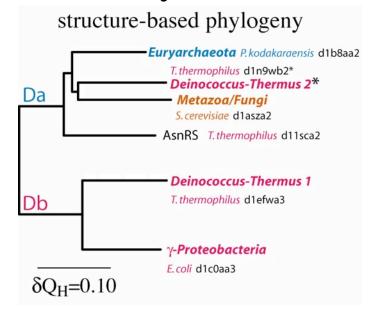
Structural Similarity Measure the effect of insertions

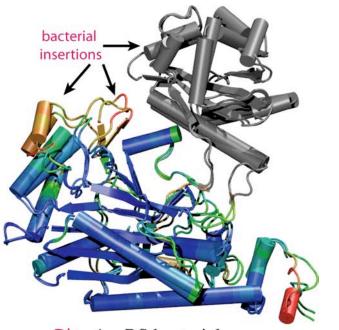
"Gaps should count as a character but not dominate" C. Woese



Protein structure encodes evolutionary information





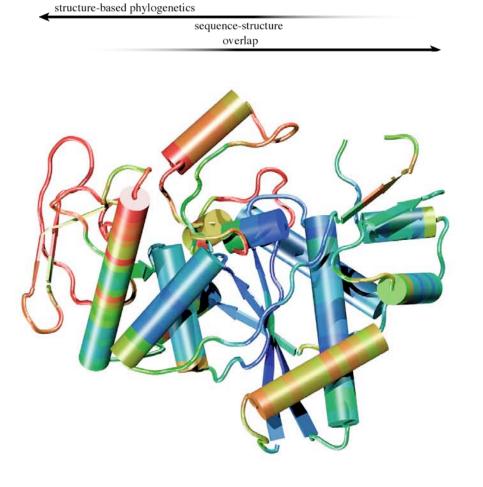


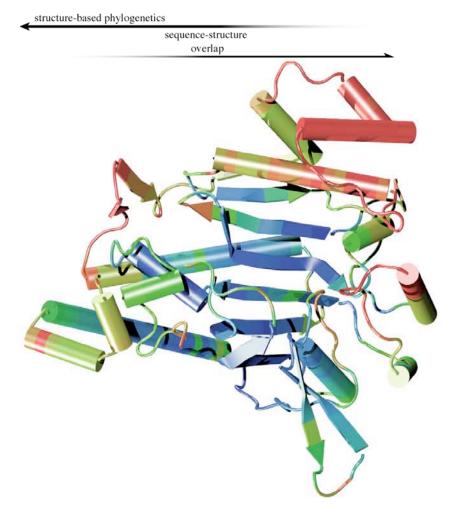
Db - AspRS bacterial genre

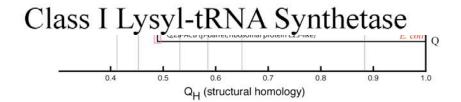
Protein structure reveals distant evolutionary events

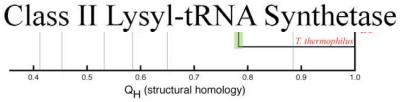
Class I AARSs

Class II AARSs









Sequences define more recent evolutionary events

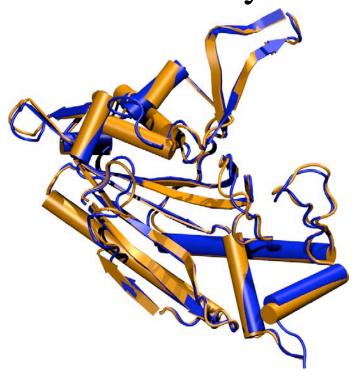


Conformational changes in the same protein.

ThrRS

T-AMP analog, 1.55 A. T, 2.00 A.

 $Q_H = 0.80$ Sequence identity = 1.00



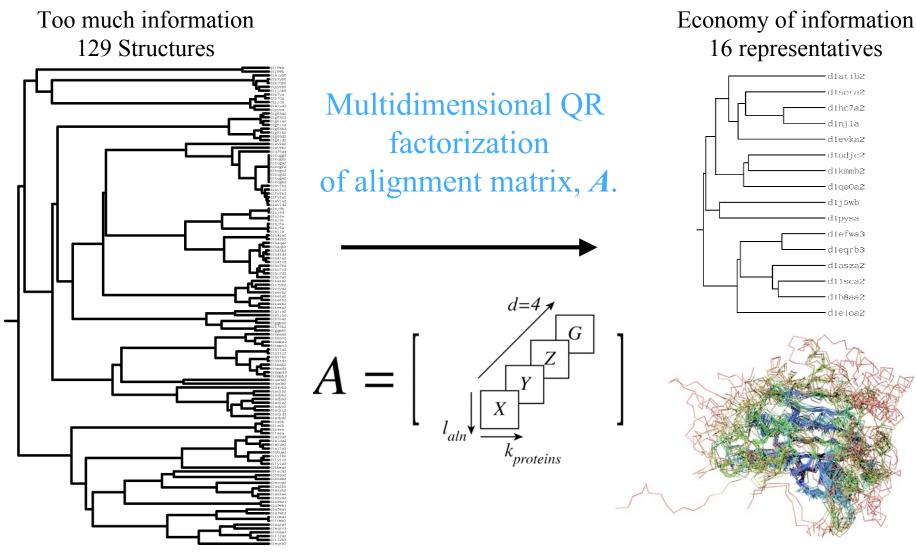
Structures for two different species.

ProRS

M. jannaschii, 2.55 A. M. thermoautotrophicus, 3.20 A.

 $Q_H = 0.89$ Sequence identity = 0.69

Non-redundant Representative Sets



QR computes a set of maximal linearly independent structures.

P. O'Donoghue and Z. Luthey-Schulten (2003) MMBR 67:550-571.

P. O'Donoghue and Z. Luthey-Schulten (2005) *J. Mol. Biol.*, **346**, 875-894.

Numerical Encoding of Proteins in a Multiple Alignment

Encoding Structure

Rotated Cartesian + Gap = 4-space

Aligned position $(x_{C_{\alpha}}, y_{C_{\alpha}}, z_{C_{\alpha}}, 0)$

Gapped position (0,0,0,g)

Gap Scaling $g = \frac{\|X\|_{F_4} + \|Y\|_{F_4} + \|Z\|_{F_4}}{\|G\|_{F_4}}$ adjustable

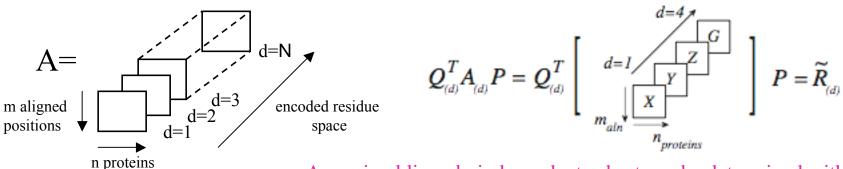
parameter

Sequence Space

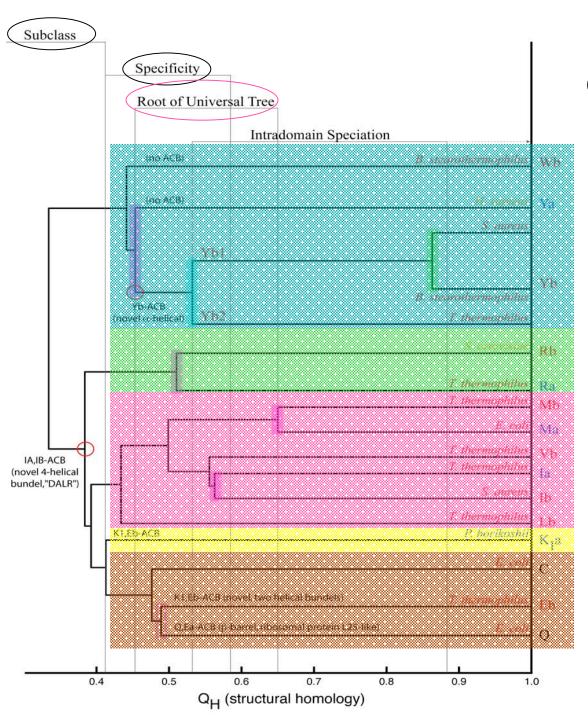
Orthogonal Encoding = 24-space

23 amino acids (20 + B, X, Z) + gap

Alignment is a Matrix with Linearly Dependent Columns



A maximal linearly independent subset can be determined with respect to a threshold, e.g., similarity measure threshold.

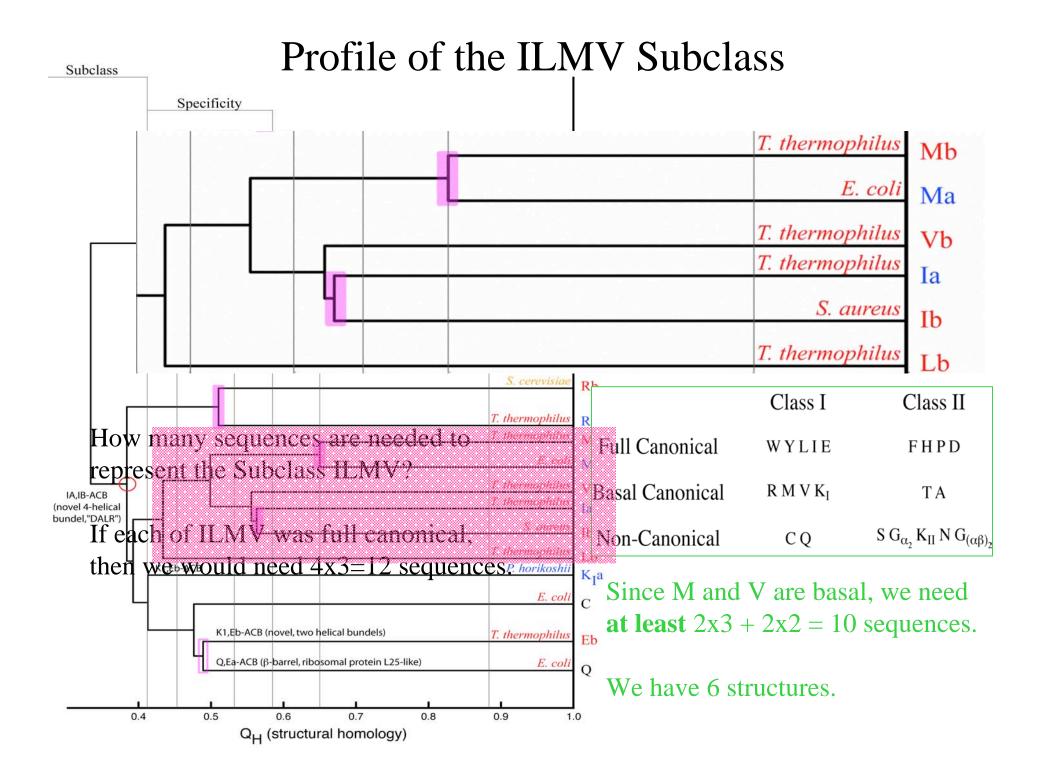


Class I AARSs evolutionary events

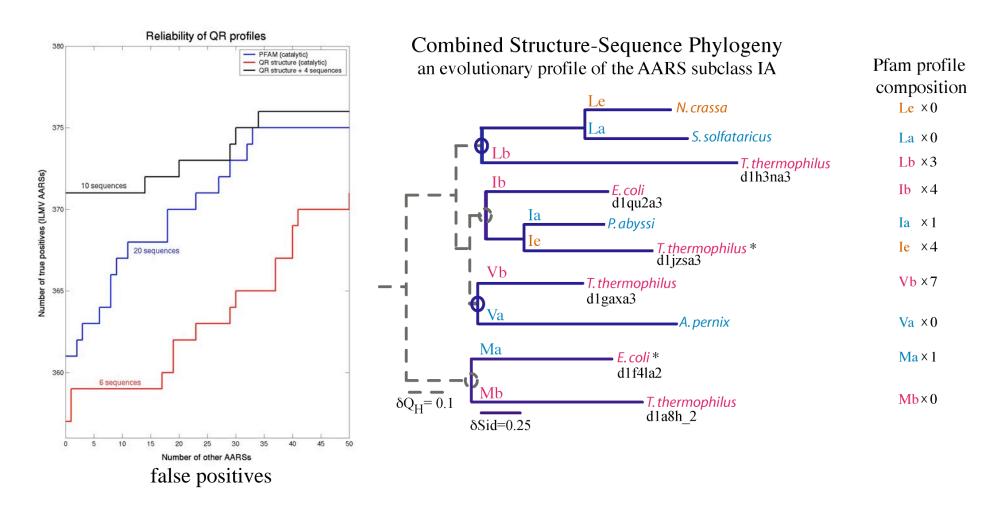
5 Subclasses

Specificity – 11 Amino acids

Domain of life A,B,E



Evolutionary Profiles for Homology Recognition AARS Subclass ILMV



The composition of the profile matters. Choosing the right 10 sequence makes all the difference.

Genome Annotation

M.jannaschii genome was completely sequenced in 1996. Genome had four missing AARSs:

```
AsnRS GlnRS Indirect Mechanism

LysRS Class I AARS

CysRS ?
```

Cysteinyl-tRNA(Cys) formation in *Methanocaldococcus jannaschii*: the mechanism is still unknown. *J. Bacteriology*, Jan. 2004, **186:**8-14. Ruan B, Nakano H, Tanaka M, Mills JA, DeVito JA, Min B, Low KB, Battista JR, and Söll D.

M. jannaschii genome database search using EP of class II AARS with HMMER

Protein	E-value	
HisRS	1.1e-10	
AspRS	1.9e-10	
PheRS α-chain	9.5e-10	
ThrRS	6.6e-04	
ProRS	9.1e-03	
SerRS	9.2e-03	
putative CysRS	1.6e-02 ← MJ166	00
AlaRS	5.1e-02	
GlyRS	0.12	
PheRS β-chain	0.15	Sethi, et. al., PNAS, 102, 2005
DNA repair protein	- 7.5	, ,

Cysteine Biosynthesis in Methanocaldococcus jannaschii

COOH
$$H \longrightarrow C \longrightarrow CH_2 \longrightarrow OPO_3H_2$$

$$ATP, tRNA^{Cys}$$

$$MJ1660 (SepRS)$$

$$MJ1594$$

$$(SerB)$$

$$COO \longrightarrow 3' tRNA^{Cys}$$

$$H \longrightarrow C \longrightarrow CH_2 \longrightarrow OPO_3H_2 + AMP + PP_i$$

$$Sep-tRNA^{Cys}$$

$$Sep-tRNA^{Cys}$$

$$Source (SepCysS)$$

$$COO \longrightarrow 3' tRNACys$$

$$SepCysS)$$

$$COO \longrightarrow 3' tRNACys$$

$$H \longrightarrow C \longrightarrow CH_2 \longrightarrow OH + PO_4^{3-}$$

$$H \longrightarrow C \longrightarrow CH_2 \longrightarrow SH + PO_4^{3-}$$

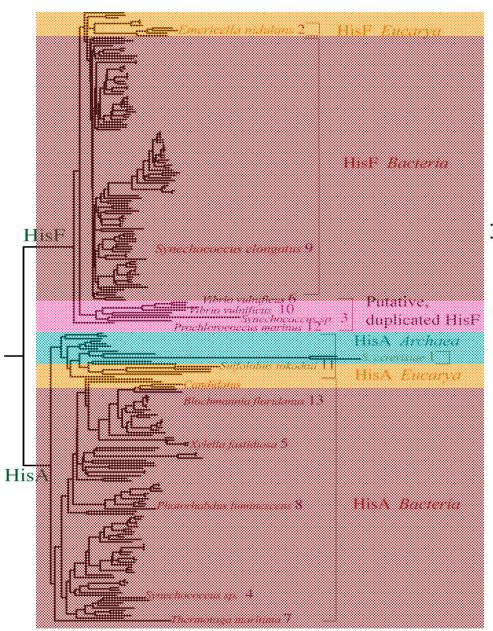
$$NH_2$$

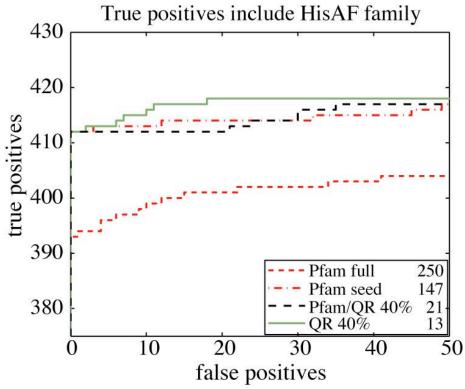
$$L-serine$$

$$Cys-tRNA^{Cys}$$

Sauerwald et al. Science 2005

Evolutionary profile for HisA-HisF family

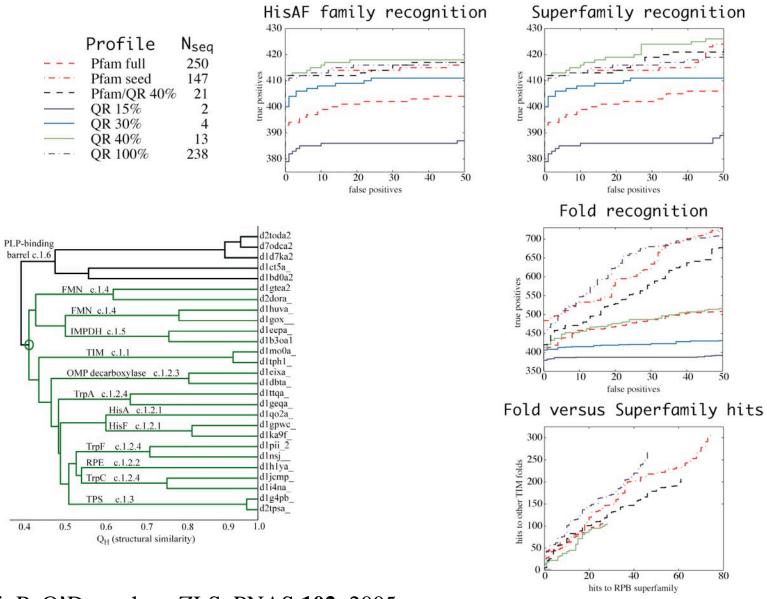




EP outperforms popular profile methods with an economy of information.

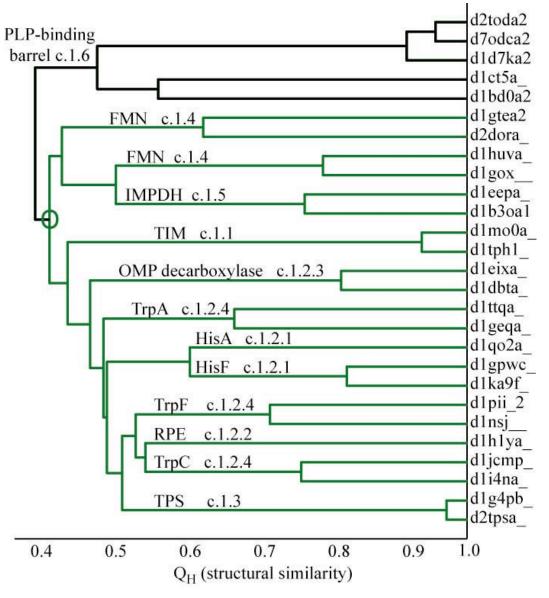
Economy of Information

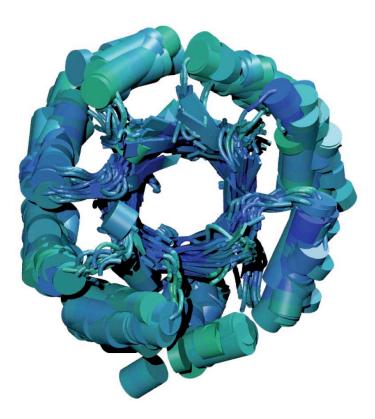
How many sequences are needed for profiles?



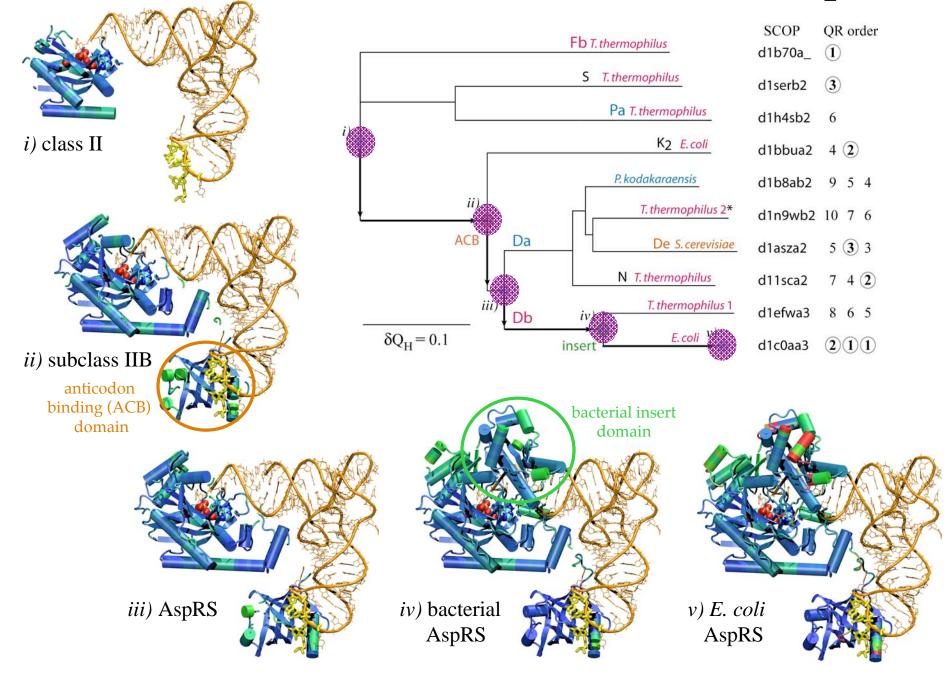
A. Sethi, P. O'Donoghue, ZLS, PNAS 102, 2005

Phylogenetic relationship between TIM barrels Found in database search with HisA-HisF profile

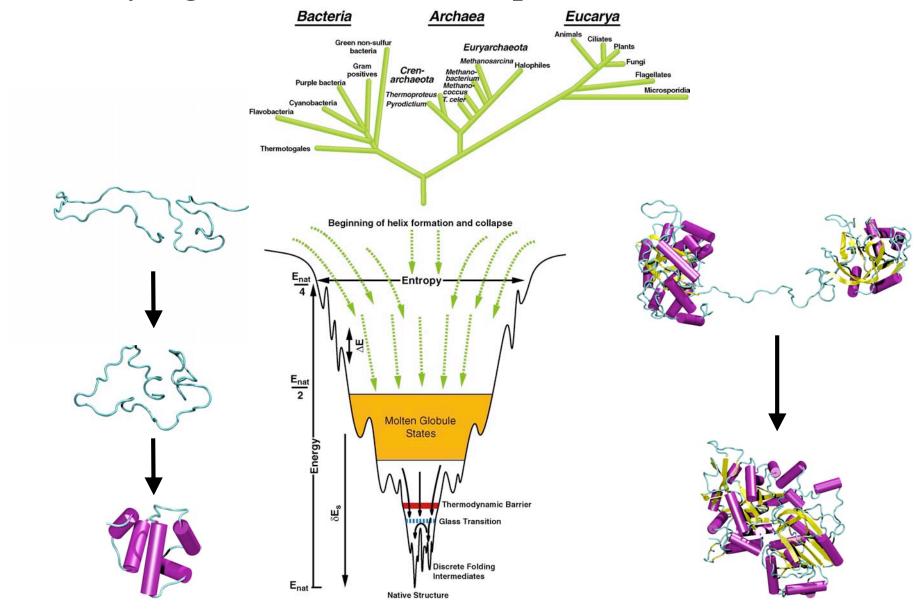




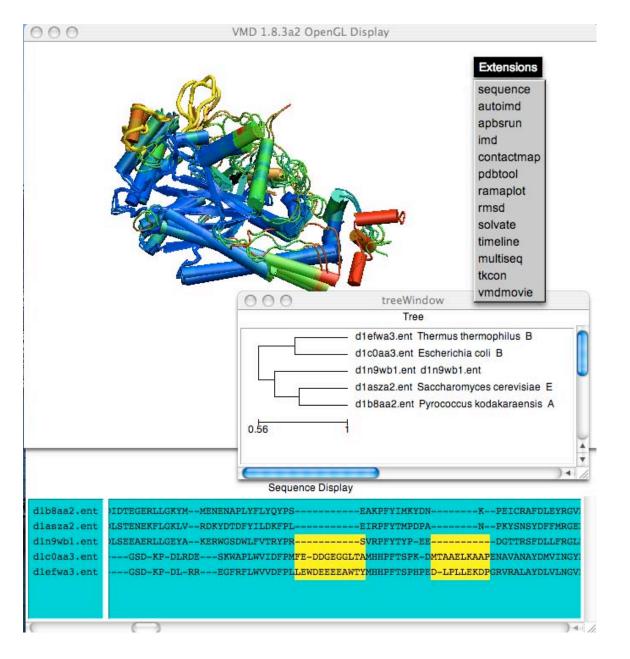
Evolution of Structure and Function in AspRS



Unifying the Worlds of Sequence and Structure



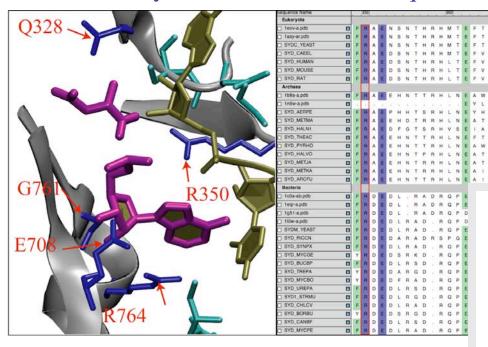
Multiseq in VMD: Merging the sequence and structure worlds



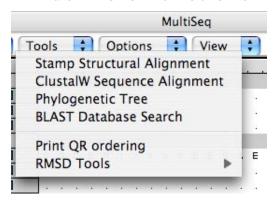
Version 1.83

2006 MultiSeq: New Features

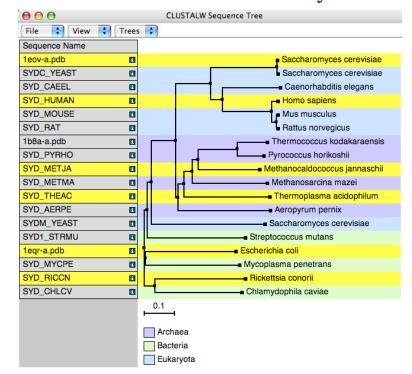
Analyze the Evolution of Sequence and Structure



Plus More Functions



Eliminate Redundancy



List of New Features in Multiseq

1. INPUT: Sequences and structures of proteins and nucleic acids from file or Blast searches of specialized databases:

Structural (PDB, SCOP, ASTRAL, NDB, VIPER..) Sequence (NCBI, ASTRAL, modified tRNA, Viral) Sequence Editor and Electronic Notebook

2. TOOLS:

Alignments (STAMP, CLUSTAL, TCoffee)
Database Searches - BLAST and VMD/Multiple DB searches
QR reduction, Phylogenetic tree - UPGMA, NJ
Conservation Mappings, RMSD plots
Covariance and Coordination Analysis

Acknowledgements

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Taras Pogorelov

Elijah Roberts

Dan Wright

Funding NSF, NIH

Graphics Programmers VMD

Elijah Roberts, Dan Wright, John Eargle
John Stone

Collaborators

Evolutionary Studies

Gary Olsen, Carl Woese (UIUC)

QR Algorithms

Mike Heath (UIUC)

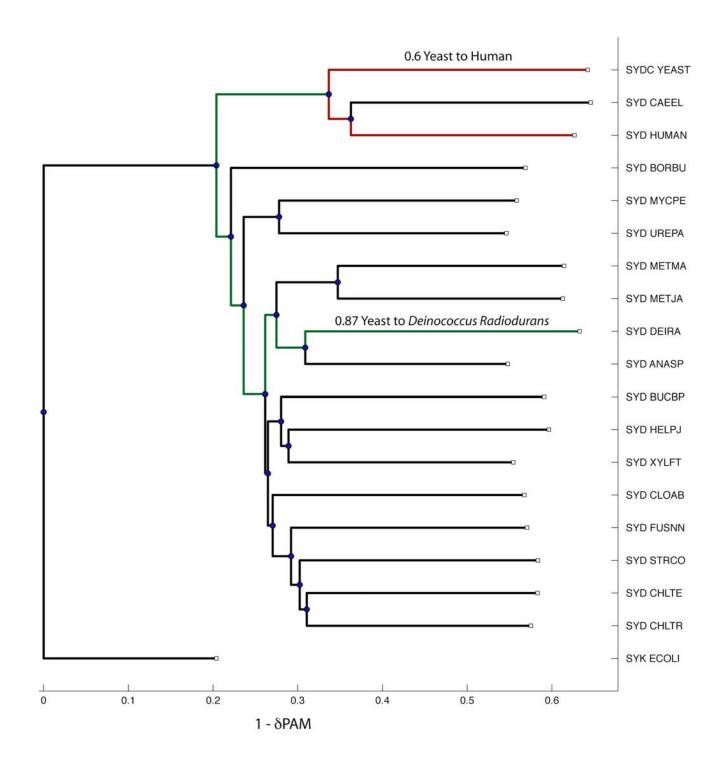
Protein Structure Prediction

Peter Wolynes, Jose Onuchic (UCSD)

Ken Suslick (UIUC)

Demonstration of New Multiseq Features

- 1. AspRS structures: STAMP multiple structure alignment. Color by structure (Qpair) and and sequence conservation. Tcl script seq ID and Sec. Str. Information in beta field.
- 2. Sequence Editor and Electronic Notebook
- 3. AspRS Sequences (from BLAST database search):
 Automated grouping by domains of life. Sequence
 conservation by domain of life. Mapping of sequence
 and structure information onto structures. CLUSTAL
 alignment to structural profile.
- 4. Phylogenetic trees of structure and sequences: HGT and QR algorithm for sequences. Evolutionary profiles

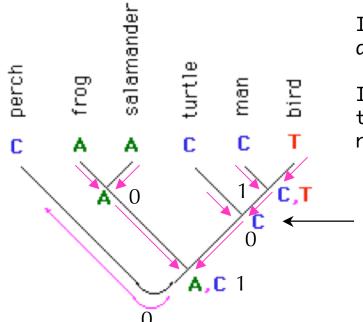


- 1. Show distance matrix for NJ/UPGMA for small number 3-4 sequences. Give algebraic equations needed for NJ.
- 2. MP/ML trees: Animate through several tree topologies generated by paup to describe the search through tree space.

Maximum Parsimony Fitch optimization

Assign characters to the ancestral nodes and calculate the number of steps (sequence changes) required by a data set on a given tree.

"Downpass" algorithm traces back through the tree from leaves to root.



If decendent characters intersect add 0 to total length.

If descendent characters do not intersect, their union set is assigned to the node add 1 to total length.

The intersection of C and (C,T) is C. This ancestral node is assigned the "state" C. The total length is unchanged.

The length on this tree for this site is 2. The length of this toplogy for the sequences in the alignment is the sum of length over all sites gives.