Supplementary Information

Effects of cytosine hydroxymethylation on DNA strand separation

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Supplementary Materials and Methods

DNA oligomers were purchased HPLC grade from IBA GmbH (Goettingen, Germany). Oligonucleotides employed had the following sequences and modifications (**hmC** represents 5-hydroxymethylcytosine):

Shear pulling geometry:

 1_{nDNA} , NH₂-(HEGL)₅-5'-(T)₁₀-CCG AGA TAT CCG CAC CAA CG-3' 2_{nDNA} , 3'-GGC TCT ATA GGC GTG GTT GC-(T)₆-5'-5'-T(Cy5)-(T)₆-GGC TCT ATA GGC GTG GTT GC-3' $1_{hmC-1-DNA}$, NH₂-(HEGL)₅-5'-(T)₁₀-CCG AGA TAT ChmCG CAC CAA CG-3' $2_{hmC-1-DNA}$, 3'-GGC TCT ATA GGhmC GTG GTT GC-(T)₆-5'-5'-T(Cy5)-(T)₆-GGC TCT ATA GGC GTG GTT GC-3' $1_{hmC-3-DNA}$, NH₂-(HEGL)₅-5'-(T)₁₀-ChmCG AGA TAT ChmCG CAC CAA hmCG-3' $2_{hmC-3-DNA}$, NH₂-(HEGL)₅-5'-(T)₁₀-ChmCG AGA TAT ChmCG CAC CAA hmCG-3' $2_{hmC-3-DNA}$, 3'-GGhmC TCT ATA GGhmC GTG GTT GhmC-(T)₆-5'-5'-T(Cy5)-(T)₆-GGC TCT ATA GGC GTG GTT GC-3' 3_{Ref} , biotin-5'-(T)₁₀-GCA ACC ACG CCT ATA GAG CC(Cy3)-3'.

Zipper pulling geometry:

 $\begin{aligned} \mathbf{1}_{nDNA}, 5'-CCG & AGA TAT CCG CAC CAA CG-(T)_{20}-(HEGL)_5-NH_2-3'\\ \mathbf{2}_{nDNA}, 5'-GGC TCT ATA GGC GTG GTT GC-(T)_6-T(Cy5)-(T)_6-CGT TGG TGC GGA TAT CTC GG-3'\\ \mathbf{1}_{hmC-1-DNA}, 5'-CCG & AGA TAT ChmCG CAC CAA CG-(T)_{20}-(HEGL)_5-NH_2-3'\\ \mathbf{2}_{hmC-1-DNA}, 5'-GGC TCT ATA GGC GTG GTT GC-(T)_6-T(Cy5)-(T)_6-CGT TGG TGhmC GGA TAT CTC GG-3'\\ \mathbf{1}_{hmC-3-DNA}, 5'-ChmCG AGA TAT ChmCG CAC CAA hmCG-(T)_{20}-(HEGL)_5-NH_2-3'\\ \mathbf{2}_{hmC-3-DNA}, 5'-GGC TCT ATA GGC GTG GTT GC-(T)_6-T(Cy5)-(T)_6-hmCGT TGG TGhmC GGA TAT CThmC GG-3'\\ \mathbf{3}_{Ref}, biotin-5'-(T)_{20}-(Cy3)-GCA ACC ACG CCT ATA GAG CC-3' \end{aligned}$

Supplementary Movies

- Movie S1 shows a trajectory (simulation C2) of hmC-3-DNA stretched in zipper geometry by steered molecular dynamics with a velocity of 1 Å/ns. The six methylated cytosines in DNA are indicated in yellow; the atoms subject to constraint (left) and stretching force (right) are shown in green.
- Movie S2 shows a trajectory (simulation F6) of hmC-3-DNA stretched in shear geometry by steered molecular dynamics with a velocity of 1 Å/ns. The methylated cytosines in DNA are indicated in yellow; the atoms subject to constraint (at bottom) and stretching force (at top) are shown in green.

Supplementary Figure



Figure S1 Time evolution of applied force (blue line) and number of base pairs (red line) as monitored in SMD simulations of unzipping nDNA (a, b), hmC-1-DNA (c, d) and hmC-3-DNA (e, f). Orange arrows indicate the breakage of hydroxymethylated CG pairs as seen by inspecting the corresponding trajectory data; black arrows indicate the breakage of normal CG pairs.



Figure S2 Snapshots of nDNA (red), hmC-1-DNA (green) and hmC-3-DNA (blue) taken at the moment when stretching force in each of the six simulations carried out for each case reaches the maximum value. The positions of hydorxymethylcytosine are marked in pink.