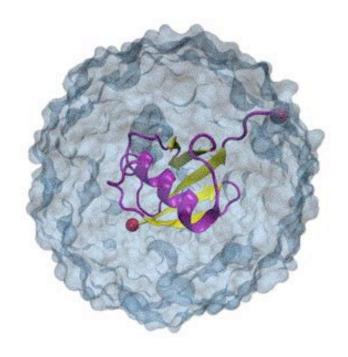
NAMD TUTORIAL

Unix/MacOSX Version



NAMD Developer: James Phillips

Timothy Isgro
James Phillips
Marcos Sotomayor
Elizabeth Villa

Scalable Molecular Dynamics with NAMD

JAMES C. PHILLIPS,¹ ROSEMARY BRAUN,¹ WEI WANG,¹ JAMES GUMBART,¹ EMAD TAJKHORSHID,¹ ELIZABETH VILLA,¹ CHRISTOPHE CHIPOT,² ROBERT D. SKEEL,³ LAXMIKANT KALÉ,³ KLAUS SCHULTEN¹

¹Beckman Institute, University of Illinois at Urbana–Champaign, Urbana, Illinois 61801 ²UMR CNRS/UHP 7565, Université Henri Poincaré, 54506 Vandœuvre-lès-Nancy, Cedex, France ³Department of Computer Science and Beckman Institute, University of Illinois at Urbana–Champaign, Urbana, Illinois 61801

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Abstract: NAMD is a parallel molecular dynamics code designed for high-performance simulation of large biomolecular systems. NAMD scales to hundreds of processors on high-end parallel platforms, as well as tens of processors on low-cost commodity clusters, and also runs on individual desktop and laptop computers. NAMD works with AMBER and CHARMM potential functions, parameters, and file formats. This article, directed to novices as well as experts, first introduces concepts and methods used in the NAMD program, describing the classical molecular dynamics force field, equations of motion, and integration methods along with the efficient electrostatics evaluation algorithms employed and temperature and pressure controls used. Features for steering the simulation across barriers and for calculating both alchemical and conformational free energy differences are presented. The motivations for and a roadmap to the internal design of NAMD, implemented in C++ and based on Charm++ parallel objects, are outlined. The factors affecting the serial and parallel performance of a simulation are discussed. Finally, typical NAMD use is illustrated with representative applications to a small, a medium, and a large biomolecular system, highlighting particular features of NAMD, for example, the Tcl scripting language. The article also provides a list of the key features of NAMD and discusses the benefits of combining NAMD with the molecular graphics/sequence analysis software VMD and the grid computing/collaboratory software BioCoRE. NAMD is distributed free of charge with source code at www.ks.uiuc.edu.

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Files needed:

```
structure mypsf.psf
coordinates mypdb.pdb
```

Define temperature

```
set temperature 310
;# target temperature used several times below
```

Starting simulation with random velocities

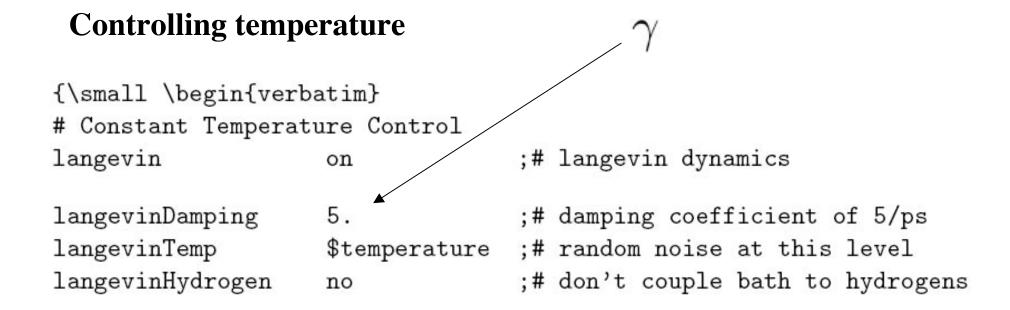
Continuing a simulation with positions and velocities from previous run

```
# continuing a run
set inputname
                   myinput
                                     ;# only need to edit this in one place!
binCoordinates
                   $inputname.coor
                                     ;# coordinates from last run (binary)
binVelocities
                   $inputname.vel
                                    ;# velocities from last run (binary)
extendedSystem
                  $inputname.xsc
                                    ;# cell dimensions from last run
                   50000
firsttimestep
                                    ;# last step of previous run
                                     ;# run stops when this step is reached
numsteps
                   100000
```

Organizing output

```
outputName
                         myoutput
            ;# base name for output from this run
                          500
                                    ;# 500 steps = every 1ps
   restartfreq
   dcdfreq
                          500
                                       Here we assume an integration time step of 2 fs,
                                       to be defined below.
   xstFreq
                          500
                                 ;# 100 steps = every 0.2 ps
outputEnergies
                       100
outputTiming
                      1000
            ;# shows time per step and time to completion
```

```
# Force-Field Parameters
    paraTypeCharmm
                        on
                        par_all27_prot_lipid.inp
    parameters
                                         12A cutoff is official standard for
    # These are specified by CHARMM
    exclude
                        scaled1-4
                                        CHARMM forcefield but smaller is
    1-4scaling
                        1.0
                                         OK when using full electrostatics
    switching
                         on
    # You have some freedom choosing the cutoff
    cutoff
                        12. ; # may use smaller, maybe 10., with PME
                        10. ;# cutoff - 2.
    switchdist
    # Promise that atom won't move more than 2A in a cycle
    pairlistdist
                        14. :# cutoff + 2.
    stepspercycle
                             ;# redo pairlists every ten steps
                        10
                                            Energy drifts if too large, but
# Integrator Parameters
                                         smaller requires more steps per ns.
                    2.0
timestep
rigidBonds
                    all
                        ;# needed for 2fs steps
nonbondedFreq
                         ;# nonbonded forces every step
                    1
fullElectFrequency 2
                         ;# PME only every other step
```



Underlying Langevin equation for all atoms

$$m_i \frac{d^2 x_i(t)}{dt^2} = F_{i,ff} - \gamma m_i \frac{dx_i(t)}{dt} + R_i(t)$$
$$\langle R_i(t) R_i(t') \rangle = 2k_B T_{\text{target}} \gamma_i \delta(t - t')$$

Using periodic boundary conditions

(avoids surface effects; permits Particle-Mesh-Ewald (PME) electrostatics; permits pressure control)

```
# Periodic Boundary conditions
cellBasisVector1
                  31.2 0. 0.; # vector to the next image
cellBasisVector2 0. 44.8 0.
cellBasisVector3 0. 0 51.3
cellOrigin
                   0. 0. ;# the *center* of the cell
wrapWater
                                 ;# wrap water to central cell
                  on
wrapAll
                                 ;# wrap other molecules too
                  on
wrapNearest
                                 ;# use for non-rectangular cells
                  off
```

Particle-Mesh-Ewald electrostatics (avoids cut-off of long-range Coulomb forces)

```
#PME (for full-system periodic electrostatics)

PME yes

PMEGridSizeX 32 ;# 2^5, close to 31.2

PMEGridSizeY 45 ;# 3^2 * 5, close to 44.8

PMEGridSizeZ 54 ;# 2 * 3^3, close to 51.3
```

Pressure Control

```
# Constant Pressure Control (variable volume)
useGroupPressure
                     yes ;# needed for rigid bonds
useFlexibleCell
                     no ;# no for water box, yes for membrane
useConstantArea
                         ;# no for water box, maybe for membrane
                     no
langevinPiston
                     on
langevinPistonTarget
                     1.01325 ;# pressure in bar -> 1 atm
                                  ;# oscillation period around 100 fs
langevinPistonPeriod
                     100.
langevinPistonDecay
                     50.
                                  ;# oscillation decay time of 50 fs
                     $temperature ;# coupled to heat bath
langevinPistonTemp
```

Underlying Langevin-Hoover barostat equation for all atoms

$$\frac{d^2V(t)}{dt^2} = -\frac{1}{W_{bs}} \left[P(t) - P_{\text{target}} \right] - \frac{1}{\tau_{\text{bs}}} \frac{dV(t)}{dt} + R_{\text{bs}}(t)$$

$$P = \rho k_B T + \frac{1}{Vd} \sum_{i < j} \langle r_{ij} \frac{dU_{tot}(r_{ij})}{dr_{ij}} \rangle \qquad d = \text{dimension}$$

$$\langle R_{\text{bs}}(t) R_{\text{bs}}(t') \rangle = \frac{2 k_B T_{\text{target}} \delta(t - t')}{W_{\text{bs}} \tau_{\text{bs}}} \qquad W_{\text{bs}} = d N_{\text{atoms}} k_B T_{\text{target}} \tau_{period}^2$$

Pressure Control

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                     50.
                                  ;# oscillation decay time of 50 fs
langevinPistonTemp
                     $temperature ;# coupled to heat bath
```

Underlying Langevin-Hoover barostat equation for all atoms: Equations solved numerically in NAMD

$$\dot{\mathbf{r}}_{i} = \mathbf{v}_{i} + s\mathbf{r}_{i} \qquad \dot{\mathbf{v}}_{i} = \mathbf{F}_{i} / m_{i} - s\mathbf{v}_{i}$$

$$\dot{V} = dVs \qquad \dot{s} = dV(P - P_{\text{target}}) / W - s / \tau_{\text{bs}} + R(t)$$

$$d - \text{dimension}$$

Fix atoms

```
fixedAtoms on
fixedAtomsFile myfixedatoms.pdb ;# flags are in this file
fixedAtomsCol B ;# set beta non-zero to fix an atom
```

Energy-minimize structure (T=0), reset temperature T, run:

```
minimize 1000 ;# lower potential energy for 1000 steps reinitvels $temperature ;# since minimization zeros velocities run 50000 ;# 100ps
```

The NAMD Output File / 1

Preamble

```
Info: NAMD 2.5b2ss03 for Linux-i686-Clustermatic
Info:
Info: Please visit http://www.ks.uiuc.edu/Research/namd/
Info: and send feedback or bug reports to namd@ks.uiuc.edu
Info:
Info: Please cite Phillips et al., J. Comp. Chem. 26: 1781-1802 (2005)
Info: in all publications reporting results obtained with NAMD.
Info:
Info: Built Fri May 30 13:09:06 CDT 2003 by jim on umbriel
Info: Sending usage information to NAMD developers via UDP.
Info: Sent data is: 1 NAMD 2.5b2ss03 Linux-i686-Clustermatic 47 umbriel jim
Info: Running on 47 processors.
```

The NAMD Output File / 2

Energies

ETITLE:	TS	BOND	ANGLE	DIHED	IMPRP
	ELECT	VDW	BOUNDARY	MISC	KINETIC
	TOTAL	TEMP	TOTAL2	TOTAL3	TEMPAVG
PRESSURE		GPRESSURE	VOLUME	PRESSAVG	GPRESSAVG
ENERGY:	1000	0.0000	0.0000	0.0000	0.0000
-97022.1848		9595.3175	0.0000	0.0000	14319.5268
-73107.3405		300.2464	-73076.6148	-73084.1411	297.7598
-6	626.5205	-636.6638	240716.1374	-616.5673	-616.6619

The NAMD Output File / 1

Writing out trajectories

•

OPENING COORDINATE DCD FILE
WRITING COORDINATES TO DCD FILE AT STEP 1000

•

Performance information

Info: Benchmark time: 47 CPUs 0.0475851 s/step 0.275377 days/ns 13540 kB memory

TIMING: 1000 CPU: 18.35, 0.01831/step Wall: 50.1581, 0.0499508/step, 6.92374 hours remaining, 14244 kB of memory in use.

Warnings

Warning: Pairlistdist is too small for 1 patches during timestep 17.

Warning: Pairlists partially disabled; reduced performance likely.

Warning: 20 pairlist warnings since previous energy output.

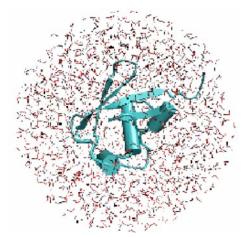
The NAMD Experience / 1

You will first simulate ubiquitin in a water sphere and water box:

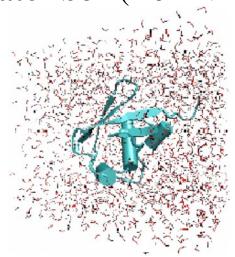
Generating a Protein Structure File (PSF)

- Go to 1-1-build directory
- Open VMD, choose extension TkCon
- Make from 1UBQ.pdb a structure without hydrogens, ubqp.pdb
- Create psf file for ubqp.pdb: ubq.pdb and ubq.psf
- Check if files exist

Solvate the protein in a water sphere (from VMD)



Solvate the protein in a water box (from VMD)



The NAMD Experience / 2

- RMSD value for equilibration
- Atomic RMSD values of equilibrated protein
- Velocity distribution
- Temperature distribution
- Specific heat
- Diffusion of whole protein
- Heat diffusion
- Temperature Echoes